2009 Vol. 11, No. 11 2465–2468

A Facile Synthetic Approach to 7-Deazaguanine Nucleosides via a Boc Protection Strategy

Ruo-Wen Wang and Barry Gold*

Department of Pharmaceutical Sciences, University of Pittsburgh, 512 Salk Hall, 3501 Terrace Street, Pittsburgh, Pennsylvania 15261 goldbi@pitt.edu

Received April 7, 2009

ABSTRACT

An efficient route to the preparation of 5-substituted 2-amino-7-((2R,4R,5R)-tetrahydro-4-hydroxy-5-(hydroxymethyl)furan-2-yl)-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one compounds has been developed by the condensation of ω -substituted aldehydes with 2,6-diaminopyrimidin-4(3*H*)-one, followed by Boc protection to afford the corresponding N^2,N^7 -tris-Boc- O^4 -t-Bu-5-substituted 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one, which is amenable to direct condensation with 1-chloro-2-deoxy-3,5-di-*O*-*p*-toluoyl- α -D-*erythro*-pentofuranose. This route affords an efficient synthesis to 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one, 2-amino-5-alkyl-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one, and guanine nucleosides.

The 5-position of 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one (aka, 7-deazaguanine) is well-suited to introduce functionalized appendages into the major groove of DNA for the purposes of structural and stability studies, DNA fluorescence labeling, DNA sequencing, and the production of DNA-based nanoarrays. There are also a number of naturally occurring 5-substituted 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one nucleosides, including Queuosine (II), and related glycosylated analogues, and Archaeosine (II)

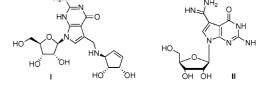


Figure 1. Queuosine (I) and Archaeosine (II).

found in tRNA (Figure 1). Accordingly, there have been numerous studies on routes to the synthesis of 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one modified nucleosides.⁷

There are several barriers to the synthesis of 5-substituted 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one nucleosides.

⁽¹⁾ Rosemeyer, H.; Ramzaeva, N.; Becker, E. M.; Feiling, E.; Seela, F. *Bioconjugate Chem.* **2002**, *13*, 1274–1285.
(2) Ju, J.; Kim, D. H.; Bi, L.; Meng, O.; Bai, X.; Li, Z.; Li, X.; Marma,

⁽²⁾ Ju, J.; Kim, D. H.; Bi, L.; Meng, Q.; Bai, X.; Li, Z.; Li, X.; Marma, M. S.; Shi, S.; Wu, J.; Edwards, J. R.; Romu, A.; Turro, N. J. *Proc. Natl. Acad. Sci. U.S.A.* **2006**, *103*, 19635–19640.

⁽³⁾ Kawate, T.; Allerson, C. R.; Wolfe, J. L. Org. Lett. 2005, 7, 3865–3868.

^{(4) (}a) Seeman, N. C. *J. Theor. Biol.* **1982**, *99*, 237–247. (b) Feldkamp, U.; Niemeyer, C. M. *Angew. Chem., Int. Ed.* **2006**, *45*, 1856–1876. (c) Seeman, N. C. *Nature* **2003**, *421*, 427–431.

⁽⁵⁾ Okada, N.; Noguchi, S.; Nishimura, S.; Ohgi, T.; Goto, T.; Crain, P. F.; McCloskey, J. A. *Nucleic Acids Res.* **1977**, *7*, 2289–2296. (b) Okada, N.; Yasuda, T.; Nishimura, S. *Nucleic Acids Res.* **1977**, *4*, 4063–4075.

⁽⁶⁾ Gregson, M.; Crain, P. F.; Edmonds, C. G.; Gupta, R.; Hashizume, T.; Phillipson, D. W.; McCloskey, J. A. *J. Biol. Chem.* **1993**, 268, 10076–10086.

^{(7) (}a) Seela, F.; Winkeler, H. D. *J. Org. Chem.* **1983**, *48*, 3119–3122. (b) Ramasamy, K.; Joshi, R. V.; Robins, R. K.; Revankar, G. R. *J. Chem. Soc., Perkin Trans. I* **1989**, 2375–2384. (c) Ramasamy, K.; Robins, R. K.; Revankar, G. R. *J. Chem. Soc., Chem. Commun.* **1989**, 560–562. (d) Seela, F.; Peng, X. *J. Org. Chem.* **2006**, *71*, 81–90. (e) Meng, Q.; Kim, D. H.; Bai, X.; Bi, L.; Turro, N. J.; Ju, J. *J. Org. Chem.* **2006**, *71*, 3248–3252. (f) Vorbrüggen, H.; Ruh-Polenz, C. *Org. React.* **2000**, *55*, 1–630.

Unlike the N1 of pyrimidines and the N9 of adenine, the N9-position of guanine, which is incorporated into a number of antiviral and anticancer nucleoside compounds, and the 5-position of 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one derivatives are not sufficiently activated for direct glycosylation. The classic solution to the preparation of 2-amino-3H-pyrrolo[2,3-d]pyrimidin-4(7H)-one nuclosides is to convert the 5-functionalized-2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7H)-one to the 4-chloro compound, which is suitable for reaction with an activated sugar (e.g., 1-chloro-2-deoxy-3,5-di-*O-p*-toluoyl-α-D-*erythro*-pentofuranose). ¹⁰ The 4-chloro nucleoside is then converted back to the keto derivative by hydrolysis. In addition to the extra synthetic steps, the 4-chloro derivatives have very poor solubility characteristics, 11 which confounds their functionalization. 7b,8b If the synthesis of 5-substituted 2-amino-3H-pyrrolo[2,3d]pyrimidin-4(7H)-one starts with 4-chloro-2-amino-3Hpyrrolo[2,3-d]pyrimidin-4(7H)-one, it must be first converted to the 5-iodo compound prior to the introduction of modifications to the 5-position by metal-mediated Sonogashira, ^{7e,12} Stille, ¹³ or related cross-coupling reactions.

In an effort to prepare a series of 5-substituted 2-amino-7-((2R,4R,5R)-tetrahydro-4-hydroxy-5-(hydroxymethyl)-fu-ran-2-yl)-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one modified DNAs (Figure 2) to (1) extend studies on how cationic and

Figure 2. 5-Aminomethyl- (left) and 5-hydroxymethyl- (right) 2-amino-7-((2*R*,4*R*,5*R*)-tetrahydro-4-hydroxy-5-(hydroxymethyl)furan-2-yl)-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-ones in DNA.

polar groups located near the floor of the major groove affect the thermodynamic stability, reactivity, and structure of DNA¹⁴ and (2) generate stable interstrand cross-links, ¹⁵ we found that the existing synthetic schemes were not suitable.

We report herein the synthesis of 5-aminomethyl- and 5-hydroxymethyl-2-amino-7-((2*R*,4*R*,5*R*)-tetrahydro-4-hydroxy-5-(hydroxymethyl)furan-2-yl)-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-ones as examples of a convenient, efficient, and general route to 5-substituted 2-amino-7-((2*R*,4*R*,5*R*)-tetrahydro-4-hydroxy-5-(hydroxymethyl)furan-2-yl)-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-ones that involves mild reaction conditions. It is also demonstrated that the approach is amenable to the preparation of guanine nucleosides.

The synthesis started with 5-substituted 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one compounds (1 and 2) that were prepared by condensation of the ω -substituted aldehydes¹⁶ with 2,6-diaminopyrimidin-4(3*H*)-one.¹⁷ As mentioned above, normally, the 5-substituted 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one would be transformed to the 4-halo derivative to activate the 7-position for reaction with a protected 1-chloro-2-deoxy- α -D-*erythro*-pentofuranose.¹⁰ Attempts to convert 1 and 2 to the 4-chloro compounds were unsuccessful.

It was envisioned that the tetra-Boc derivatives of 1 and 2 could be prepared and then selectively deprotected to reveal the pyrrole NH-7 for coupling with a reactive Cl sugar (Figure 3).

Figure 3. Design of 5-substituted 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one as new coupling precusors with improved solubility.

Compounds 1 and 2 have very limited solubility, so they were treated as a suspension in MeCN with excess Boc₂O. After several days at rt, all of the solid starting material had gone into solution. Instead of the anticipated tetra-Boc derivative, it was found that 1 and 2 afforded the tris-Boc-protected *O*⁴-*t*-Bu ether compounds 7 and 8, respectively (Scheme 1). Fortuitously, formation of the *O*-*t*-Bu ethers negates the need to convert the 2-amino-3*H*-pyrrolo[2,3-*d*]pyrimidin-4(7*H*)-one nucleus into the 4-chloro derivative prior to sugar coupling.

To elucidate the origin of the O^4 -t-Bu ethers 7 and 8, the reactions were monitored by TLC and LC/MS analysis. Time

2466 Org. Lett., Vol. 11, No. 11, 2009

^{(8) (}a) Sims, K. A.; Woodland, A. M. *Pharmacotherapy* **2006**, *12*, 1745–1757. (b) Field, A. K.; Tuomari, A. V.; McGeever-Rubin, B.; Terry, B. J.; Mazina, K. E.; Haffey, M. L.; Hagen, M. E.; Clark, J. M.; Braitman, A.; Slusarchyk, W. A. *Antiviral Res.* **1990**, *13*, 41–52. (c) Brigden, D.; Fiddian, P.; Rosling, A. E.; Ravenscroft, T. *Antiviral Res.* **1981**, *1*, 203–212. (d) Vere Hodge, R. A.; Sutton, D.; Boyd, M. R.; Harnden, M. R.; Jarvest, R. L. *Antimicrob. Agents Chemother.* **1989**, *33*, 1765–7173. (e) Lee, J.; Chuang, T.-H.; Redecke, V.; She, L.; Pitha, P. M.; Carson, D. A.; Raz, E.; Cottam, H. B. *Proc. Natl. Acad. Sci. U.S.A.* **2003**, *100*, 6646–6651.

^{(9) (}a) Tolman, R. L.; Tolman, G. L.; Robins, R. K.; Townsend, L. B. *J. Heterocycl. Chem.* **1970**, *7*, 799–806. (b) Seela, F.; Westermann, B.; Bindig, U. *J. Chem. Soc., Perkin Trans. I* **1988**, 697–702.

⁽¹⁰⁾ Ramasamy, K.; Imamura, N.; Robins, R. K.; Revankar, G. R. *Tetrahedron Lett.* **1987**, *43*, 5107–5100.

⁽¹¹⁾ Dey, S.; Garner, P. J. Org. Chem. 2000, 65, 7697–7699.

^{(12) (}a) Jager, S.; Rasched, G.; Kornreich-Leshem, H.; Engeser, M.; Thum, O.; Famulok, M. J. Am. Chem. Soc. **2005**, 127, 15071–15082. (b) Seela, F.; Shaikh, K. I. Tetrahedron **2005**, 61, 2675–2681.

⁽¹³⁾ Angelov, T.; Guainazzi, A.; Scharer, O. D. Org. Lett. 2009, 11, 661-664.

^{(14) (}a) Manning, G. S. *Q. Rev. Biophys.* **1978**, 2, 179–246. (b) Record, M. T.; Anderson, C. F.; Lohman, T. M. *Q. Rev. Biophys.* **1978**, 2, 103–179. (c) Honig, B.; Nicholls, A. *Science* **1995**, 268, 1144–1149. (d) Gold, B. *Biopolymers* **2002**, 65, 173–179. (e) Gold, B.; Marky, L. M.; Stone, M. P.; Williams, L. D. *Chem. Res. Toxicol.* **2006**, 19, 1402–1414.

^{(15) (}a) Wilds, C. J.; Noronha, A. M.; Robidoux, S.; Miller, P. S. *Nucleosides Nucleotides Nucleic Acids* **2005**, *4*, 965–969. (b) Räschle, M.; Knipsheer, P.; Enoiu, M.; Angelov, T.; Sun, J.; Griffith, J. D.; Ellenberger, T. E.; Schärer, O. D.; Walter, J. C. *Cell* **2008**, *134*, 969–980.

⁽¹⁶⁾ De Luca, L.; Giacomelli, G.; Porcheddu, A. Org. Lett. 2001, 3, 3041–3043,

^{(17) (}a) Klepper, F.; Polborn, K.; Carell, T. *Helv. Chim. Acta* **2005**, *88*, 2610–2616. (b) Barnett, C. J.; Grubb, L. M. *Tetrahedron Lett.* **2000**, *41*, 9741–9745.

Scheme 1. Preparation of O-t-Bu Ethers

R O Boc₂O, DMAP TEA, MeCN NBoc₂

1:
$$X = C$$
, $R = CH_2NPhth$
2: $X = C$, $R = CH_2OBn$
7-deazaguanine: $X = C$, $R = H$
guanine: $X = N$

R O NBoc₂

7: $X = C$, $R = CH_2NPhth$
8: $X = C$, $R = CH_2NPhth$
9: $X = C$, $R = CH_2OBn$
9: $X = C$, $R = H$
10: $X = N$

course studies clearly showed the buildup of the tetra-Boc derivatives (presumably $\bf 3$ and $\bf 4$) and subsequent loss of CO_2 with ether formation to give $\bf 7$ and $\bf 8$, respectively (Figure 4).

Figure 4. Products 7 and 8 formed from the reactions of 1 and 2 with t-Boc₂O, respectively, that arise via intermediates 3 and 4.

In terms of the scope of the reaction, the tetra-Boc intermediate and the butyl ether product were also observed in the reaction of unsubstituted 2-amino-3*H*-pyrrolo[2,3-d]pyrimidin-4(7*H*)-one, indicating that the functionality attached to the 5-position does not play a role in the conversion of the carbamate to the ether (9). The reaction with guanine was explored despite previous reports on the difficulty of protecting guanine with Boc₂O. ¹⁰ After 96 h, the N^2 , N^2 , N^7 -tris-Boc- O^4 -t-Bu ether derivative of guanine was isolated and characterized. Earlier time points in the reaction were analyzed by LC/MS and clearly showed that formation of ether product (10) proceeds through the initial formation of the tetra-Boc derivative.

The conversion of **7** to the protected 2'-deoxynucleoside is shown in Scheme 2. To reveal the pyrrole NH-7, the N^7 -Boc was selectively deprotected by sodium methoxide to give pyrrolo[2,3-d]pyrimidinone **12**, which is an efficient precursor, with good solubility, for the sugar coupling reaction. Compound **12** was condensed with 1-chloro-2-deoxy-3,5-di-O-p-toluoyl- α -D-erythro-pentofuranose (**30**) to give the desired β -anomer of the protected 2'-deoxynucleoside **13**. The phthalimide and Boc protecting groups were sequentially removed with hydrazine and TFA, and the primary amine was then protected as the trifluoroacetamide **14**. From compound **7**, compound **14** was prepared in five steps in 42% yield, and only one column purification was required.

Scheme 2. Synthesis of 16

The toluoyl groups were selectively removed with $Mg(OMe)_2$ to give trifluoroacetamide **15**. The N^2 -amino group in **15** was selectively protected to give isobutyric amide **16**. The overall yield from **1** to **16** is 6.2%.

Compound **16** was converted into the *O3'*-(2-cyanoet-hoxy)(diisopropylamino)phosphino]-5'-*O*-(4,4'-dimethoxytrityl) derivative **18** (Scheme 3) by standard procedures to

Scheme 3. Syntheses of Phosphamidite 18

provide the required intermediate for solid phase DNA synthesis. 19

Similarly, the conversion of **8** to the deoxynucleoside **27** involved its coupling to the chlorosugar after selective removal of the N^7 -Boc group with NaOMe (Scheme 4).

Org. Lett., Vol. 11, No. 11, 2009

⁽¹⁸⁾ Xu, Y.-C.; Lebeau, E.; Walker, C. Tetrahedron Lett. 1994, 35, 6207–6210.

^{(19) (}a) BeaucageS. L. In *Protocols for Oligonucleotides and Analogs*; Agrawal, S., Ed.; Humana Press: Totowa, NJ, 1993; pp 33–62. (b) Bleasdale, B.; Ellwood, S. B.; Golding, B. T. *J. Chem. Soc., Perkin Trans. I* **1990**, 803–805.

Scheme 4. Syntheses of Compound 27

The O3'- and O5'-toluoyl protection was converted to TBDMS protection by sequential treatment with NaOMe and TBSMSCl. Reaction with Boc₂O restored the bis- N^2 -Boc protection, and the O-Bn group was then reductively removed by Pd(OH)₂-catalyzed hydrogenation. The free primary alcohol **23** was reprotected as the benzoate **24**, and the N^2 -

Boc and O^4 -t-butyl groups were removed by heating at 80 °C in vacuo on silica. The N^2 -position was reprotected with i-PrCOCl and the silyl protection removed with TBAF. The overall yield of **27** from **2** is 6.4% via 14 steps. The primary alcohol **23** can, if desired, be further reduced to give 5-methyl-2-amino-3H-pyrrolo[2,3-d]pyrimidin-4(7H)-one, a useful isosteric analogue of 7-methylguanine. The Bz group of ester **24** can also be removed in acidic conditions used for removal of N-Boc groups. The synthesis of compound **27** illustrated that Boc protection strategy provides an efficient route to the preparation of those 5-substituted 2-amino-7-((2R,4R,5R)-tetrahydro-4-hydroxy-5-(hydroxymethyl)furan-2-yl)-3H-pyrrolo[2,3-d]pyrimidin-4(7H)-one with senstive functionalities since Boc and O^4 -t-butyl groups can be efficiently removed under mild condition.

In conclusion, we have synthesized 5-aminomethyl-2-amino-7-((2R,4R,5R)-tetrahydro-4-hydroxy-5-(hydroxymethyl)furan-2-yl)-3H-pyrrolo[2,3-d]pyrimidin-4(7H)-one (**16**) and 5-hydroxymethyl-2-amino-7-((2R,4R,5R))-tetrahydro-4-hydroxy-5-(hydroxymethyl)furan-2-yl)-3H-pyrrolo[2,3-d]pyrimidin-4(7H)-one (**27**) from **1** and **2**, respectively, in 6% yield via key O^4 -t-Bu ether intermediates **7** and **8**. These syntheses illustrate an efficient and general route to the preparation of 5-substituted 2-amino-7-((2R,4R,5R))-tetrahydro-4-hydroxy-5-(hydroxymethyl)furan-2-yl)-3H-pyrrolo[2,3-d]pyrimidin-4(7H)-ones. Time course studies showed that the tetra-Boc derivatives were converted to the O^4 -t-Bu ethers via intramolecular transformation.

Acknowledgment. This work was supported by NIH RO1 CA29088.

Supporting Information Available: Experimental procedure and spectral data for synthesized compounds. This material is available free of charge via the Internet at http://pubs.acs.org.

OL9007537

(20) Apelqvist, T.; Wensbo, D. Tetrahedron Lett. 1996, 37, 1471-1472.

2468 Org. Lett., Vol. 11, No. 11, 2009